

An asymmetric synthesis of ADDA and ADDA-glycine dipeptide using the β -lactam synthon method

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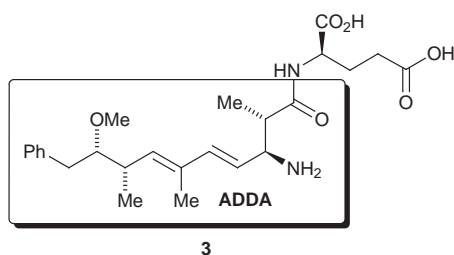
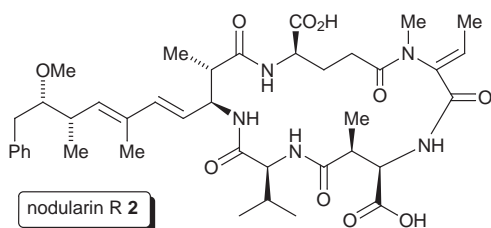
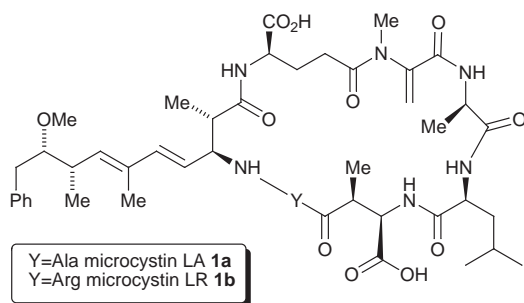
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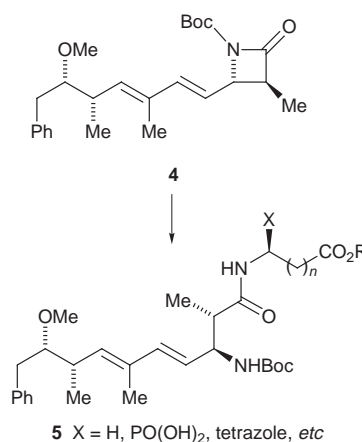
The paper describes the synthesis of the *N*-Boc lactam **4** and demonstrates that it is an important intermediate in the synthesis of dipeptide **5** ($X = H, n = 0, R = Me$), an analogue of the ADDA-Glu dipeptide **3**. In addition we have described a mild method for the preparation of the amino acid salt ADDA·HCl and provided synthetic methods and full characterisation for the previously 'elusive' free amino acid ADDA.

Introduction

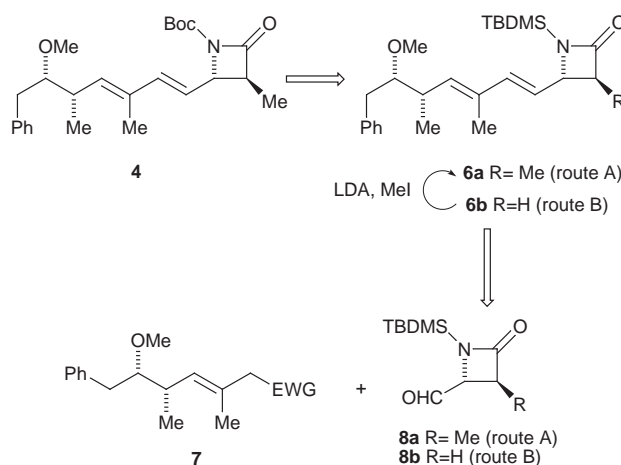
The microcystin (*e.g.*, microcystin LA **1a** and LR **1b**) and nodularin (*e.g.*, nodularin R **2**) families of cyclic peptides are potent hepatotoxins.² In order to probe their structure-activity relationship we wanted to prepare analogues of the ADDA-glutamic acid dipeptide **3**, a common feature of these peptides. As *N*-Boc



β -lactams have recently been shown to give dipeptides on reaction with amino esters,³ we identified the *N*-Boc β -lactam **4** as a versatile intermediate to achieve this goal (Scheme 1). This paper describes the synthesis of the β -lactam **4**, its reaction with glycine methyl ester to form an analogue **5** ($X = H, n = 0, R = Me$) of the dipeptide **3**, and in addition provides the first full characterisation of the parent amino acid, ADDA.



Scheme 1



Scheme 2

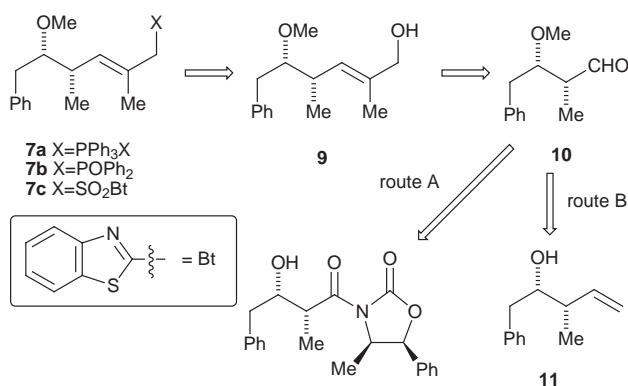
The retrosynthesis of β -lactam **4** is outlined in Scheme 2. The β -lactam ring provides both a means for the preparation of the required peptide bond of compound **5** and a scaffold for the introduction of the *anti*- α -methyl- β -amino acid motif within the ADDA amino acid residue as C-4-substituted β -lactams are known to undergo enolate alkylations at C-3 to give predominantly the *trans*-disubstituted products.⁴ Due to the high reactivity of *N*-Boc lactams,^{3,5} we considered the *N*-Boc lactam **4** too unstable to participate in the required enolate alkylation chem-

istry. However, as *N*-TBDMS β -lactams are known to undergo enolate alkylations and the *N*-TBDMS bond is easily cleaved,^{4,6} our initial target became lactam **6a** or **6b**. We chose to assemble the diene fragment within lactam **6a** or **6b** through condensation of an allylic coupling partner **7** and β -lactam aldehyde **8a** or **8b**.^{6b} While the introduction of the C-2 methyl group could take place either before (route A, Scheme 2) or after (route B, Scheme 2) the diene assembly, the more convergent route A became the focus of our initial investigations, the result of which are described here.⁷

Results and discussion

Synthesis of allylic coupling partners **7a–c**

The synthesis of the phosphonium salt **7a** has been described previously using the general retrosynthesis illustrated in Scheme 3, route A and applied to the synthesis of ADDA derivatives.^{8a–c,f} These literature reactions using the phosphonium salt **7a** give approximately a 1:1 mixture of *E* and *Z* isomers, so while the phosphonium salt **7a** was to be assessed in this work, we were also interested in investigating the coupling using the phosphine oxide **7b**⁹ and the benzothiazole sulfone **7c**^{10,†} due to their efficacy in the synthesis of related *E,E*-dienes.^{11,12} The common intermediate for compounds **7a–c** is the allylic alcohol **9**, prepared from the aldehyde **10** by a Wittig reaction/reduction procedure. Of the two standard methods for the stereospecific preparation of aldehyde **10**, we chose the Brown crotylation methodology¹³ (Scheme 3, route B) as

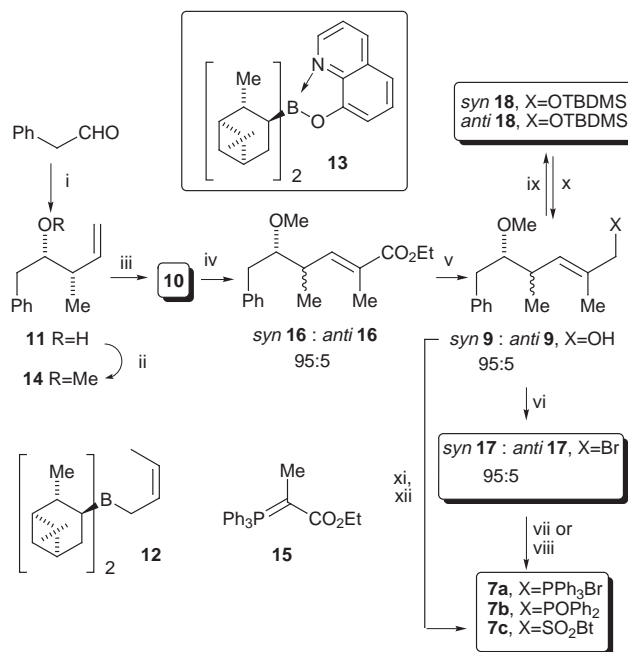
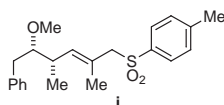


Scheme 3

opposed to the previously described Evans methodology^{8c,f,14} due to the fact that (i) the terminal alkene is a more convenient source of the aldehyde than an Evans's oxazolidone, (ii) the methyl ether could be introduced through standard Williamson synthesis without competing retro-aldol fragmentation, and (iii) the intermediates could be purified by distillation, allowing the synthesis of compounds **7a–c** to be more conveniently carried out on a large scale.

Reaction of phenylacetaldehyde with the borane **12**¹³ and work-up with 8-hydroxyquinoline^{13c} gave the *syn*-homoallylic alcohol **11** (Scheme 4). ¹H NMR analysis confirmed both the diastereo- and enantioselectivity for this reaction to be >95%.¹⁵ The work-up procedure permitted the pinene-derived chiral auxiliary to be removed as the solid complex **13**, allowing the alcohol **11** to be isolated in the filtrate of the work-up solution. The alcohol **11** was directly methylated to give the alkene

† The "traditional" sulfone **i** has been applied to the synthesis of ADDA derivatives (ref. 8d,e).



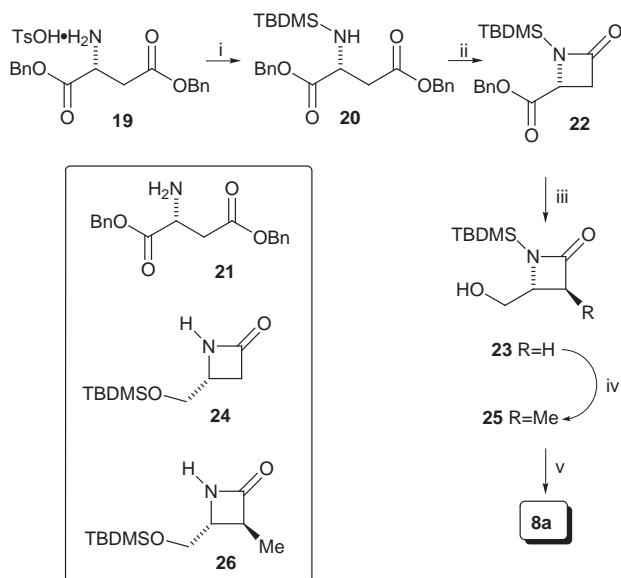
Scheme 4 Reagents: (i) **12**; (ii) NaH, MeI; (iii) O₃, PPh₃; (v) **15**; (v) LiAlH₄; (vi) CBr₄, PPh₃; (vii) PPh₃, *anti* **7a**:*syn* **7b**, 95:5; (viii) EtOPPh₂, recryst., *anti* **7b**:*syn* **7b**, 100:0; (ix) TBDMSCl; (x) MeOH, acid; (xi) BtSH, DEAD, PPh₃; (xii) Oxone or H₂O₂, *anti* **7c**:*syn* **7c**, 100:0.

14. The efficacy of the Brown crotylation combined with the 8-hydroxyquinoline work-up and simple short-path distillation of the alkene **14** allowed the conversion of phenylacetaldehyde to the alkene **14** to be conducted on a 80 mmol scale in 85–90% yield. Ozonolysis of the alkene **14** followed by reductive work-up gave the aldehyde **10**^{8b–e,g} which was treated without purification with the Wittig reagent **15** to give, after short-path distillation, the $\alpha\beta$ -unsaturated ester **16** in 80–85% yield. ¹H NMR analysis indicated that the *E*:*Z* ratio was >20:1, but that the expected product *syn*-**16**^{8b–e} was contaminated with ~5% of the C-4 epimer *anti*-**16**, which could not be removed by chromatography. A similar level of epimerisation has been reported previously for this reaction.^{8b–e} Reaction of the aldehyde **10** with the Horner–Wadsworth–Emmons reagent, triethyl 2-phosphonopropionate, favoured the formation of *Z*-**16** (ratio of *Z*-**16**:*E*-**16**, 60:40) in addition to inducing a similar level (~5%) of epimerisation. LiAlH₄ reduction^{8d,e} of the mixture of esters *syn*-**16** and *anti*-**16** (ratio 95:5) followed by work-up with Na₂SO₄·10H₂O gave a mixture of alcohols *syn*-**9**^{8a,f} and *anti*-**9** in quantitative yield. Conversion of this mixture of alcohols to the bromides *syn*-**17**^{8b,c,f} and *anti*-**17** (65–70% yield) and reaction with PPh₃ gave the phosphonium salts *syn*-**7a** and *anti*-**7a**.^{8a–c,f} As attempted recrystallisation of this material was unsuccessful, the phosphonium salt was used in subsequent reactions as the 95:5 mixture of *syn* and *anti* diastereoisomers. Alternatively, reaction of the bromides *syn*-**17** and *anti*-**17** with EtOPPh₂ gave the crude phosphine oxide **7b**. Recrystallisation gave diastereoisomerically pure phosphine oxide *syn*-**7b** in 70–75% yield. The allylic alcohol *syn*-**9** could be separated from stereoisomer *anti*-**9** after conversion to the TBDMS ethers *syn*-**18** and *anti*-**18**.^{8e} While the *R_f* difference between isomers *syn*-**18** and *anti*-**18** was small, significant quantities of *syn*-**18** could be purified and converted to alcohol *syn*-**9**. Mitsunobu reaction of alcohol *syn*-**9** and 2-mercaptobenzothiazole (BtSH) followed by oxidation of the intermediate sulfide with either Oxone[®]¹⁶ or molybdenum-catalysed H₂O₂¹² gave the sulfone *syn*-**7c** in 70–75% yield for the two steps. Direct recrystallisation of the sulfone **7c** prepared from the mixture of alcohols *syn*-**9** and *anti*-**9** did not remove all of the unwanted *anti*-isomer, although the recrystallisation procedure has not been repeated after a

seed crystal of diastereoisomerically pure sulfone *syn-7c* was prepared from alcohol *syn-9*.

Synthesis of β -lactam aldehyde **8a**

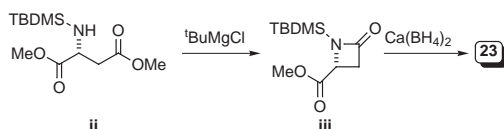
The synthesis of aldehyde **8a** is shown in Scheme 5. The



Scheme 5 Reagents: (i) TBDMSCl, NEt₃; (ii) ^tBuMgCl; (iii) NaBH₄, LiBr; (iv) base, MeI (see text); (v) oxidation (see text).

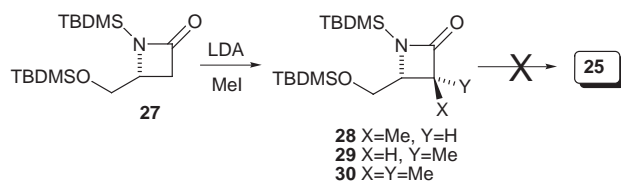
toluene-*p*-sulfonic acid salt of dibenzyl-*D*-aspartate¹⁷ **19** was treated with TBDMSCl in the presence of 2 equivalents of Et₃N under anhydrous conditions to give dibenzyl *N*-TBDMS-*D*-aspartate **20** in >90% yield. Compound **20** is sufficiently stable in solution to allow the liberated Et₃N·HCl to be removed by aqueous work-up; however, the neat liquid was relatively unstable and prolonged storage or extensive manipulation led to decomposition to give the free amine **21** and therefore it was used in further reactions immediately. *ent-20* has previously been prepared in a two-step process from the salt *ent-19* by liberation of the free amine *ent-21* and subsequent reaction with *N*-(*tert*-butyldimethylsilyl)-*N*-methyltrifluoroacetamide.^{4f} Reaction of compound **20** with ^tBuMgCl proceeded smoothly to give the benzyl ester **22** in addition to an equivalent of benzyl alcohol.^{4f} This crude material was treated with NaBH₄ in the presence of LiBr to give the alcohol **23** and a further equivalent of benzyl alcohol.¹⁸ The alcohol **23** could be easily separated from the two equivalents of benzyl alcohol by silica gel chromatography. Ca(BH₄)₂ has been recommended⁴ⁿ for the reduction of ester (\pm)-**22** and related compounds in order to suppress migration of the TBDMS group from nitrogen to oxygen; however, use of the more convenient NaBH₄/LiBr reduction procedure led to only trace amounts of the lactam **24**,^{4d} which was removed during chromatography. Reaction of 30–70 mmol of salt **19** delivered a 50–60% yield of alcohol **23** in a 3-step process requiring only the final product to be purified.‡ Reaction of the alcohol **23** with two equivalents of

‡ The *N*-silylation/cyclisation of the dimethyl ester **ii** to give the lactam **iii** was less efficient (~40% yield) when compared to the formation of lactam **22** from diester **20**. The conversion of **ii** to **iii** is described (no yield or characterisation) in a patent (ref. 19) outlining the synthesis of compound **27**. Curiously, reduction of ether **iii** to alcohol **23** with NaBH₄/LiBr gave a mixture of compounds **23** and **24**, whereas Ca(BH₄)₂ reduction of compound **iii** gave only alcohol **23** (see ref. 4n).



n-BuLi or LDA at –78 °C followed by addition of methyl iodide (1–3 equiv.) and quenching the reaction at –78 °C gave the *trans*-methylated lactam **25** stereospecifically as shown by the H-2–H-3 coupling of 2.4 Hz.^{4g,n} As was expected no evidence was seen in the crude ¹H NMR spectrum of the *cis*-methylated lactam. Despite the reaction not proceeding beyond 70% conversion, the starting material could be easily separated from the product by chromatography and recycled. The isolated yield of the *trans*-methylated product **25** was 53–66% on a multigram scale. Varying the base [LDA, *n*-BuLi, lithium hexamethyldisilazide (LiHMDS)], the reaction time (both in the formation of the dianion and the MeI quench), the ratio of reagents or the addition of adjuncts (*e.g.* HMPA) all failed to improve the conversion beyond 70%. Attempts to perform the reaction at temperatures greater than –78 °C resulted in increasing amounts of the lactams **24** and **26** being isolated as a result of N → O silyl migration. At –30 °C the sole products of the alkylation were those derived from N → O silyl migration. Treatment of the alcohol **23** with either Dess–Martin's reagent²⁰ or under Swern²¹ oxidation conditions gave the aldehyde **8a** in 90% yield. Due to the large molecular mass of Dess–Martin's reagent, the Swern protocol was preferred for larger scale oxidations. The aldehyde retained the *trans*-substitution of the methyl and formyl groups about the lactam ring (*J*_{3,4} 2.8 Hz). The crude aldehyde **8a** obtained from the Swern oxidation procedure was of sufficient purity to be used immediately. Material of greater purity for characterisation was obtained by silica gel chromatography. In general, the aldehyde **8a** could be stored unchanged for weeks at –10 °C; however, as on one occasion some decomposition occurred, the aldehyde **8a** was used immediately it was prepared.

Thomas and Williams have reported^{4k,§} that the methylation of the bis-TBDMS lactam **27** gave the mono-*trans*-alkylated lactam **28** in addition to ~10% of another compound, assumed to be the *cis*-lactam **29** (Scheme 6). Significantly, no bis-methyl-

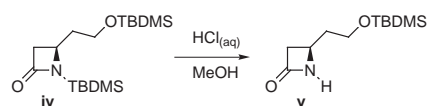


Scheme 6

ated lactam **30** was detected. Indeed, lactam **28** was resistant to further alkylation using LDA as the base. In an effort to increase the amount of available alcohol **25** and hence aldehyde **8a**, we attempted the preparation of the methylated lactam **28** followed by selective removal of the *O*-TBDMS group.¶ Reaction of compound **27** with a large excess of LDA and MeI, under the conditions reported by Thomas and Williams, gave a quantitative yield of a mixture of the desired monomethyl lactam **28** and, surprisingly, the 3,3-dimethyl lactam **30** in a ratio of 3:1. A further reaction using a slight excess of LDA and MeI gave an improved ratio of products **28**:**30** of 9:1; however, no further improvements could be made and attempts to separate the two products by chromatography proved fruitless. Other workers have recently reported that dimethylation occurs during large-scale reactions of lactam *ent-27* with excess of LDA and MeI.²³ In addition, the *O*-TBDMS group could not be

§ Thomas *et al.* performed the alkylation experiments on the opposite enantiomer, *i.e.*, lactam **27** derived from (*L*)-aspartic acid.

¶ A patent abstract (ref. 22) describes the selective *N*-desilylation of lactam **iv** (and other *N*,*O*-silyl lactams) to give compound **v**.

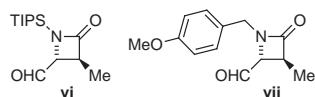


removed without some *N*-TBDMS cleavage so this route was abandoned.

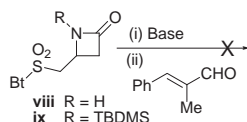
Coupling of the allylic reagents 7a–c with β -lactam aldehyde 8a

The couplings of 7a–c and the aldehyde 8a were studied extensively (Scheme 7). Deprotonation of the phosphonium salt 7a with *n*-BuLi and addition of aldehyde 8a in THF gave the lactam 6a in 25% yield with an *E*:*Z* ratio of ~1:1. No attempt was made to optimise this reaction as we investigated more stereoselective processes. Deprotonation of the phosphine oxide 7b with NaHMDS and addition of the aldehyde 8a gave only the *E*-isomer; however, the yield of lactam 6a was poor (10–15%). While some phosphine oxide 7b could be recovered from these reactions, no aldehyde 8a was detected in the crude reaction mixture even in reactions using >2 equivalents of aldehyde 8a. The only isolated and identified product derived from aldehyde 8a (other than diene 6a) was *tert*-butyldimethylsilylanol. The exact mode of decomposition of aldehyde 8a remains unclear at this stage. Sodium seems to be the optimum counter-ion in this reaction, as replacement of NaHMDS by *n*-BuLi or KHMDS gave no identifiable products in the crude reaction mixture. || Curiously, inverse addition of the sodium salt of the phosphine oxide 7b to a solution of aldehyde 8a also gave no identifiable products. Replacing the phosphine oxide 7b by the sulfone 7c in the coupling reaction led to a significant increase in yield of coupled product. The yield and *E*:*Z* ratio of the diene from these reactions in THF varied with the nature of the base used to deprotonate the sulfone. To aid purification, the crude products were treated with KF to give the lactam 31 as the isolated product. Examination of the products isolated in this fashion indicated that KHMDS was the base of choice, delivering lactam 31 with a ~3:1 *E*:*Z* ratio in 40–45% yield over the two steps (coupling and desilylation) from sulfone 7c. Reactions using NaHMDS were highly *E*-selective (*E*:*Z* ratio ~4:1); however, the yield was inferior (20–25%). Use of LiHMDS or LDA as base gave the poorest *E*:*Z* ratio of ~2:1. Reaction of the lactam 31 with (Boc)₂O gave the *N*-Boc lactam 4 in 90–95% yield, which on reaction with glycine methyl ester in the presence of NaN₃^{3,24} gave the ADDA-Glu analogue dipeptide 5 (X = H, *n* = 0, R = Me) in 76% yield, significantly without epimerisation. The identity of the *N*-Boc lactam 4 was confirmed by conversion to *N*-Boc-ADDA^{8c,f,h,i} using Greico's pro-

|| Poor yields of coupled products were also obtained on reaction of phosphine oxide 7b with aldehyde 8b. While again only the desired *E*,*E*-diene was obtained, other products derived from substrate 8b could not be isolated or identified. It should be noted that the reaction of aldehyde 8b with CBr₄/Zn^{6c} or Reformatsky,^{6a} Peterson^{4j} and stabilized Wittig^{4k,l} reagents is reported to give the desired products in good yield. Reaction of the triisopropylsilyl (TIPS) lactam vi gave no coupled product and led to the destruction of substrate vi. Reaction of the *N*-*p*-methoxybenzyl (PMB) lactam vii with phosphine oxide 7b gave an improved (30%) isolated yield of coupled product but in this case a ~1:1 mixture of inseparable *E* and *Z* diene isomers was obtained. In addition, oxidative deprotection with cerium(IV) ammonium nitrate (CAN) of this mixture led to decomposition.



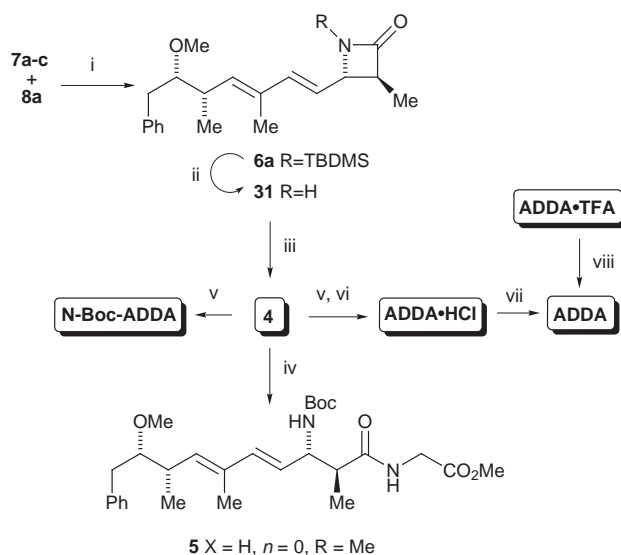
Reversing the polarity of the coupling process was briefly investigated. Attempted double deprotonation of compound (±)-viii or selective deprotonation of compound (±)-ix followed by addition of α -methylcinnamaldehyde gave no diene products.



cedure.²⁵ Samples of the *N*-Boc lactam 4 containing mixtures of *E*- and *Z*-isomers could be separated by column chromatography either at this stage or after coupling to form the dipeptide 5. This reaction confirms that the *N*-Boc lactam 4 is a useful intermediate for the synthesis of dipeptides 5. Future work in this area includes further optimising the synthesis of the *N*-Boc lactam 4 and applying it to the synthesis of particular ADDA-containing dipeptides 5 as required for the synthesis of 'designer' microcystins and nodularins.

Synthesis of ADDA

While there have been several papers over recent years⁸ detailing synthetic approaches to various derivatives of ADDA, there is only one report^{8a} of the preparation of ADDA although no experimental details were given and only an accurate mass measurement was reported. The lack of spectral data is in part due to the instability of ADDA under the conditions used to degrade microcystins and nodularins to their constituent amino acids.^{8a} Full characterisation of a compound assigned as the free amino acid ADDA appeared in a recent thesis,^{8c} but as the last purification step in this procedure was an extraction into dil. HCl, it appears this compound was the hydrochloride salt. In order to provide quantities of ADDA for a variety of applications, we were interested in developing methods for the conversion of our synthetic intermediates to ADDA. As acid hydrolysis has been shown to be an effective method for the conversion of β -lactams to β -amino acids,³ we treated β -lactams 6 and 31 under a variety of acidic conditions, *e.g.* 6 M HCl in CHCl₃, 6 M HCl in MeOH, TMSCl in MeOH; however, no identifiable products were isolated. It appears that conditions vigorous enough to effect ring opening of the lactam also cause decomposition. The successful mild basic hydrolysis of the *N*-Boc lactam 4 to give *N*-Boc-ADDA described above led us to consider this intermediate as a precursor to ADDA (Scheme 7). To this end, *N*-Boc-ADDA was treated with HCl in



Scheme 7 Reagents (i) see text; (ii) KF, MeOH; (iii) (Boc)₂O; (iv) Gly(OMe), NaN₃; (v) LiOH; (vi) HCl, EtOAc; (vii) HCO₂NH₄ (aq); (viii) NH₃ (aq).

EtOAc²⁶ under mild conditions to give ADDA as its HCl salt in ~95% purity as assessed by HPLC analysis. The ¹H and ¹³C NMR spectral data for this material agreed with those previously prepared and reported as being for the free amino acid ADDA.^{8c} Purification of this material by HPLC (elution with 6:4 methanol–water containing ammonium formate) surprisingly gave the free amino acid ADDA. Apparently exchange of chloride ion for formate ion occurs on the column to give the formate salt of ADDA which, under high vacuum, decomposes to the free amino acid ADDA which was fully characterised. In

an alternative procedure, ADDA·TFA** was dissolved in aq. ammonia and the solution freeze dried to again give the free amino acid ADDA. In this case, ADDA is exchanged for ammonia to give ammonium trifluoroacetate which decomposes to trifluoroacetic acid and ammonia under high vacuum. Examination of the ¹H NMR spectra for the free amino acid ADDA showed that the resonances for H-2 and H-3 appear as broad pseudo-triplets with coupling of 7.2 and 8.4 Hz respectively. No other coupling is observed. In addition the chemical shifts for H-2 and H-3 were 0.2–0.4 ppm upfield of the respective peaks in the spectra for either ADDA·HCl or ADDA·TFA.

In conclusion, this work has described the synthesis of the *N*-Boc lactam **4** and demonstrated that it is an important intermediate in the synthesis of ADDA compound **5** (X = H, *n* = 0, R = Me), an analogue of the ADDA-Glu dipeptide **3**. In addition we have described a mild method for the preparation of the amino acid salt ADDA·HCl and provided synthetic methods and full characterisation for the previously 'elusive' free amino acid ADDA.

Experimental

General

Mps were determined using a Kyoma hotstage melting point apparatus and are uncorrected. Short-path distillations were performed using a Kugelrohr (bulb-to-bulb) distillation apparatus and temperatures are oven temperatures and serve only as a guide. Microanalysis were performed at the Chemistry Department, University of Otago, New Zealand. Optical rotations were measured using a JASCO DIP370 digital polarimeter in a cell of 1 dm in length at a wavelength of 598 nm (sodium D-line). Concentrations are expressed as *c* (g/100 ml). The temperature of all rotations was 22 ± 1 °C. [*α*]_D-Values are given in units of 10⁻¹ deg cm² g⁻¹. IR spectra were recorded using a Perkin-Elmer 842 spectrometer (cm⁻¹ scale) for samples as KBr disks of solids or as thin films of liquids between sodium chloride plates. ¹H NMR spectra were recorded at 200 MHz with a Bruker AC-200 spectrometer and refer to deuteriochloroform solutions with residual chloroform as the internal standard (*δ*_H 7.27) unless otherwise stated. *J*-Values are given in Hz. ¹³C NMR spectra were recorded at 50 MHz with a Bruker AC-200 spectrometer and refer to deuteriochloroform solutions with residual chloroform as the internal standard (*δ*_C 77.0). ¹⁹F NMR spectra were recorded at 188 MHz with a Bruker AC-200 spectrometer and refer to deuteriochloroform solutions with chemical shifts reported relative to CCl₃F (*δ* 0.00). Low-resolution CI MS and accurate mass determinations were recorded on a JOEL JMS-DX303 mass spectrometer. Atmospheric pressure chemical ionisation (APCI) MS were recorded on a FISIONS Instrument VG Platform mass spectrometer. Analytical HPLC was performed on a LiChroCart C18 RP-HPLC column, 4 mm I.D. × 125 mm. All solvents were purified by literature procedures.²⁷ Unless otherwise stated, all reagents were purchased from Aldrich Chemical Company, Inc. Petroleum spirit refers to the fraction with distillation range 40–60 °C.

(3*S*,4*S*)-4-Methoxy-3-methyl-5-phenylpent-1-ene **14**

(*Z*)-But-2-ene (23.2 ml, 250 mmol) was condensed into a solution of potassium *tert*-butoxide in THF (1 M; 91.6 ml, 91.6 mmol) at -78 °C. *n*-BuLi in hexanes (2.5 M; 18.3 ml, 91.6 mmol) was added dropwise while the internal temperature was maintained at below -70 °C. The reaction temperature was then allowed to reach -45 °C for 10 min and the mixture was recooled to -78 °C. (+)-β-Methoxydiisopinocampheylborane (32.9 g, 104

mmol) as a solution in Et₂O (60 ml) was added dropwise at -78 °C and the solution was stirred at this temperature for 45 min. Freshly distilled BF₃·OEt₂ (15.8 ml, 129 mmol) was added dropwise followed immediately by a solution of phenylacetaldehyde (10.0 g, 83.2 mmol) in Et₂O (40 ml) and the mixture was stirred at -78 °C overnight. The reaction was quenched with addition of anhydrous MeOH (16.8 ml, 415 mmol) and the mixture was concentrated *in vacuo* to leave a viscous oil. This residue was dissolved in anhydrous MeOH (150 ml), and the solution was cooled to 0 °C and treated with a solution of 8-hydroxyquinoline (15.1 g, 104 mmol) in MeOH (150 ml) to give a fluorescent yellow-green solution. The mixture was stirred overnight during which it reached ambient temperature and a fluorescent yellow-green solid had precipitated. Filtration through a short column of Florisil[®], washing of the filter cake with small quantities of cold MeOH, and concentration of the filtrate *in vacuo* gave a fluorescent liquid. ¹H NMR analysis indicated the mixture contained (3*S*,4*S*)-4-hydroxy-3-methyl-5-phenylpent-1-ene **11** [¹H NMR: *δ* 1.13 (d, *J* 6.8, 3H), 2.27–2.40 (m, 1H), 2.61 (dd, *J* 9.3 and 13.7, 1H), 2.85 (dd, *J* 3.7 and 13.7, 1H), 3.68–3.77 (m, 1H), 5.08–5.10 (m, 1H), 5.14–5.19 (m, 1H), 5.76–5.96 (m, 1H), 7.19–7.37 (m, 5H) in addition to ~10% of borinate **13**. The enantiomeric excess of alcohol **11** was determined to be >95% after conversion to the corresponding (*R*)- and (*S*)-Mosher's esters [¹⁹F NMR: *δ* -71.76 {major peak using (*R*)-Mosher's acid}, -72.01 {major peak using (*S*)-mosher's acid}].

This material was treated further without purification. NaH (60% in oil; 5.0 g, 124.8 mmol) was added in portions to a solution of the crude alcohol in THF (200 ml) containing methyl iodide (10.4 ml, 166.4 mmol) at 0 °C. The cooling bath was removed and the mixture was stirred until TLC analysis indicated the reaction was complete. The reaction mixture was cooled to 0 °C and treated cautiously with saturated aq. NH₄Cl (20 ml) and diluted with Et₂O (200 ml) and water (200 ml). The organic layer was separated, dried (MgSO₄), and filtered and the filtrate was concentrated *in vacuo* to give a yellow oil. Short-path distillation (150 °C at 0.1 mmHg) gave the *methyl ether* **14** (11.8 g, 75%) as a slightly yellow oil (Found: C, 81.8; H, 9.7. C₁₃H₁₈O requires C, 82.1; H, 9.5%); [*α*] -26.5 (*c* 1, CHCl₃); *v*_{max}(film) 1641m and 1605m cm⁻¹; ¹H NMR *δ* 1.17 (d, *J* 7.0, 3H), 2.31–2.47 (m, 1H), 2.70 (dd, *J* 7.9 and 10.4, 1H), 2.82 (dd, *J* 4.6 and 10.4, 1H), 3.26 (s, 3H), 3.23–3.30 (m, 1H), 5.02–5.06 (m, 1H), 5.09–5.12 (m, 1H), 5.81–5.99 (m, 1H) and 7.21–7.31 (m, 5H); ¹³C NMR *δ* 15.2, 37.9, 40.9, 58.4, 86.5, 114.6, 126.0, 128.3, 129.4, 139.8 and 141.2; *m/z* (CI) 191 (M⁺ + 1, 100%), 135 (70) and 91 (40).

(2*E*,4*S*,5*S*)-Ethyl 5-methoxy-2,4-dimethyl-6-phenylhex-2-enoate *syn*-**16**

O₃ in O₂ was bubbled through a solution of alkene **14** (6.0 g, 31.6 mmol) in CH₂Cl₂ (200 ml) at -78 °C. The clear solution turned blue after *ca.* 90 min. The solution was degassed with N₂ for 5 min, triphenylphosphine (9.11 g, 34.8 mmol) was added, and the reaction mixture was allowed to warm to rt and was stirred for 2 h. While routinely this CH₂Cl₂ solution of aldehyde **10** was used directly in the next reaction, on occasions the solution was concentrated *in vacuo* to give neat aldehyde **10**, whose spectra agreed with those previously reported.^{8c,g,j} (Ethoxycarbonylethylidene)triphenylphosphorane **15** (22.7 g, 63.2 mmol) was added and the mixture was heated to reflux for 48 h. The reaction mixture was allowed to cool to ambient temperature and was concentrated *in vacuo*. The residue was dissolved in CH₂Cl₂ (10 ml) and applied to the top of a silica gel pad and eluted with 50% Et₂O in petroleum spirit (500 ml). The filtrate was concentrated *in vacuo* and purified by short-path distillation (150 °C at 0.05 mmHg) to give a clear oil (7.3 g). ¹H NMR analysis indicated the products of the reaction were the *ester syn*-**16** (diagnostic peak 3-H, *δ* 6.69, dq, *J* 1.5 and 10.2)

** ADDA·TFA was prepared by Dr Raghu Samy of Professor Peter Toogood's laboratory at the Department of Chemistry at the University of Michigan. Peter Toogood is currently at Parke Davis Pharmaceutical Research, Ann Arbor, Michigan.

and the corresponding 4*R* epimer **anti-16** (diagnostic peak 3-H, δ 6.81, dq, *J* 1.5 and 9.9) in a ratio of 95:5. No evidence of the (2*Z*,4*S*,5*S*)-(diagnostic peak 3-H, δ 5.96, dq, *J* 1.5 and 9.7) or (2*Z*,4*R*,5*S*)-(diagnostic peak 3-H, δ 5.86, dq, *J* 1.5 and 9.9) isomers was detected. The spectral data for the ester **syn-16** agreed with those reported previously^{8c,e} and the product from this reaction was used without additional purification.

(2*E*,4*S*,5*S*)-5-Methoxy-2,4-dimethyl-6-phenylhex-2-en-1-ol **syn-9**

LiAlH₄ (1.0 M in Et₂O; 39.6 ml, 39.6 mmol) was added dropwise to a solution of the esters **syn-16** and **anti-16** (7.3 g, 26.4 mmol) in Et₂O (200 ml) at -78 °C. The reaction mixture was stirred at this temperature for 1 h after which time the solid CO₂-acetone-bath was replaced with an ice-bath and the mixture was stirred for a further 1 h. TLC analysis [silica gel; 25% Et₂O in petroleum spirit] indicated no starting material (*R*_f 0.8) remained. Na₂SO₄·10H₂O was added while the internal temperature was maintained below +10 °C. Aq. NaOH (1 ml; 30%) was added and the mixture was stirred for 30 min. Anhydrous NaSO₄ was added, the reaction mixture was filtered, and the filtrate was concentrated *in vacuo* to give the alcohol **syn-9**, containing 5% of **anti-9**, as a clear oil (6.2 g, 100%). The spectral data for the alcohol **syn-9** agreed with those reported previously^{8c,e} and the product from this reaction was used without additional purification.

(2*E*,4*S*,5*S*)-5-Methoxy-2,4-dimethyl-6-phenylhex-2-enyl-(triphenyl)phosphonium bromide **7a**

CBr₄ (16.9 g, 51 mmol) in CH₃CN (100 ml) was added dropwise to a degassed solution of the mixture of alcohols **syn-9** and **anti-9** prepared above (6.2 g, 26.5 mmol) and PPh₃ (13.4, 51 mmol) in CH₃CN (200 ml) at rt in the dark. Cooling of the reaction mixture to 0 °C during the addition of the CBr₄ solution resulted in precipitation of PPh₃. While reactions conducted without degassing of the mixture and exposed to light gave substantial bromide, some oxidation of the alcohol to the corresponding aldehyde occurred. The mixture was stirred overnight at rt and was then concentrated *in vacuo*. Column chromatography (silica gel; petroleum spirit) gave the desired bromide containing ~30% CHBr₃. Short-path distillation (130 °C at 0.07 mmHg) gave the (2*E*,4*S*,5*S*)-1-bromo-2,4-dimethyl-5-methoxy-6-phenylhex-2-ene **syn-17**, containing 5% of **anti-17**, as clear oil (5.4 g, 68%) whose spectral data agreed with those previously reported for this compound.^{8c} The mixture of bromides was treated with PPh₃ according to the general procedure outlined in ref. 8c to give the phosphonium salt **7a** as a solid which could not be purified by recrystallisation and was used directly in subsequent coupling reactions. ¹H NMR (major **syn-7a** isomer only) δ 0.80 (d, *J* 7.0, 3H), 1.39 (dd, *J* 1.3 and 3.3, 3H), 2.15–2.26 (m, 1H), 2.44–2.53 (m, 2H), 3.02–3.11 (m, 1H), 3.12 (s, 3H), 4.44–4.58 (m, 1H), 4.79–4.93 (m, 1H), 5.31–5.39 (m, 1H), 7.03–7.34 (m) and 7.61–7.94 (m, ArH over-integrates due to small amounts of triphenylphosphine).

(2*E*,4*S*,5*S*)-1-Diphenylphosphinoyl-5-methoxy-2,4-dimethyl-6-phenylhex-2-ene **syn-7b**

The mixture of bromides **syn-17** and **anti-17** prepared using the method described above (5.4 g, 18.0 mmol) was dissolved in THF (50 ml) and treated with a solution of ethyl diphenylphosphinite (4.2 g, 18.2 mmol) in THF (50 ml). The solution was degassed with Ar and heated to reflux overnight. Concentration of the reaction mixture *in vacuo* and recrystallisation of the residue from 50% Et₂O in petroleum spirit (60 ml) gave the phosphine oxide **syn-7b** (5.6 g, 51% from the 95:5 mixture of alcohols **syn-9** and **anti-9**), mp 87–88 °C (Found: C, 77.2; H, 7.5. C₂₇H₃₁O₂P requires C, 77.5; H, 7.5%); [α] -10.0 (*c* 1, CHCl₃); ν_{\max} (KBr) 2948s, 1436s, 1184s, 1105s, 734s, 699s and 555s cm⁻¹; ¹H NMR δ 0.80 (d, *J* 6.8, 3H), 1.65 (dd, *J* 1.3 and

2.7, 1H), 2.29–2.49 (m, 2H), 2.62 (dd, *J* 4.6 and 13.9, 2H), 2.93–3.03 (m, 1H), 3.05–3.14 (m, 2H), 3.10 (s, 3H), 4.97–5.05 (m, 1H), 6.98–7.34 (m, 5H), 7.31–7.62 (m, 6H) and 7.66–8.08 (m, 4H); ¹³C NMR δ 16.0 (*J* 3.6), 18.4, 36.5 (d, *J* 2.2), 37.8, 41.0 (d, *J* 68), 58.3, 86.5 (d, *J* 2.1), 125.7 (d, *J* 10.2), 125.7, 127.9, 128.4 (dd, *J* 5.8 and 11.6), 129.3, 130.8–131.0 (m), 131.4–131.6 (m), 132.0 (d, *J* 11.4) and 133.8 (d, *J* 10.1); *m/z* (APCI) 419 (*M*⁺ + 1, 100%) and 117 (30).

2-[(2*E*,4*S*,5*S*)-5-Methoxy-2,4-dimethyl-6-phenylhex-2-enyl]-sulfonylbenzothiazole **syn-7c**

The alcohol **syn-9** was separated from alcohol **anti-9** after conversion to the TBDMS ethers **syn-18** and **anti-18** according to ref. 8e. DEAD (4.25 ml, 27.0 mmol) was added dropwise to a solution of the alcohol **syn-9** (5.75 g, 24.6 mmol), BtSH (4.51 g, 27.0 mmol) and triphenylphosphine (7.08 g, 27.0 mmol) in THF (300 ml) at -10 °C. The reaction mixture was stirred for 1 h at this temperature and concentrated *in vacuo*. Column chromatography (silica gel; 30% Et₂O in petroleum spirit) gave 2-[(2*E*,4*S*,5*S*)-5-methoxy-2,4-dimethyl-6-phenylhex-2-enyl]-thio}benzothiazole as clear oil (9.4 g, 100%), [α] -43.1 (*c* 1, CHCl₃); ν_{\max} (film): 2932s, 2826s, 1603w, 1459s and 1428 cm⁻¹; ¹H NMR δ 0.97 (d, *J* 6.8, 3H), 1.67 (d, *J* 1.3, 1H), 2.35–2.53 (m, 1H), 2.57 (dd, *J* 7.4 and 13.9, 1H), 2.73 (dd, *J* 4.6 and 13.9, 1H), 3.07–3.15 (m, 1H), 3.17 (s, 3H), 3.99 (ABX, *J* 0.9 and 13.3, 2H), 5.52 (dq, *J* 1.2 and 9.7, 1H), 7.06–7.45 (m, 7H), 7.71–7.76 (m, 1H) and 7.87–7.92 (m, 1H). ¹³C NMR δ 15.3, 15.8, 36.5, 37.7, 42.8, 58.3, 86.4, 120.6, 121.2, 123.9, 125.6, 125.7, 127.8, 129.0, 129.1, 133.2, 134.9, 138.9, 152.8 and 166.4; *m/z* (CI) 384 (*M*⁺ + 1, 65%), 185 (30), 168 (95), 135 (100) and 134 (40). This material was oxidised directly using one of the following procedures.

Method A. Oxone[®] (3.34 g, 10.98 mmol of KHSO₅) in water (40 ml) was added dropwise to a solution of the sulfide (1.40 g, 3.66 mmol) in MeOH (40 ml) with the internal temperature kept below +10 °C. The cooling bath was removed and the mixture was stirred at rt and monitored by TLC. The sulfide (*R*_f 0.9; silica gel; 30% Et₂O in petroleum spirit) was converted to the more polar (*R*_f 0.3) sulfoxide within 1 h, which was itself oxidised more slowly to the desired sulfone (*R*_f 0.45) after 6 h at rt. The mixture was diluted with water, extracted with CHCl₃ (3 × 100 ml) and the organic extracts were dried (MgSO₄), filtered, and concentrated *in vacuo* to give a clear oil. Purification by column chromatography (silica gel; 30–70% Et₂O in petroleum spirit) gave the sulfone **7c** (1.10 g, 72%) as a solid, mp 88.5–89 °C (Found: C, 63.5; H, 6.2; N, 3.5. C₂₂H₂₅NO₃S requires C, 63.6; H, 6.1; N, 3.4%); [α] -25.1 (*c* 1, CHCl₃); ν_{\max} (KBr) 2929s, 2894s, 1661s and 1474s cm⁻¹; ¹H NMR δ 0.80 (d, *J* 7.0, 3H), 1.65 (d, *J* 1.3, 1H), 2.34–2.59 (m, 3H), 2.88–2.96 (m, 1H), 3.02 (s, 3H), 4.16 (s, 2H), 5.26 (d, *J* 9.9, 1H), 7.00–7.26 (m, 5H), 7.48–7.64 (m, 2H), 7.91–7.96 (m, 1H) and 8.18–8.23 (m, 1H). ¹³C NMR δ 14.9, 16.6, 36.3, 37.4, 58.0, 64.1, 85.7, 121.6, 121.9, 125.0, 125.7, 127.3, 127.6, 127.8, 128.9, 136.5, 138.6, 140.3, 152.3 and 165.2; *m/z* (APCI) 416 (*M*⁺ + 1, 100%), 384 (50) and 136 (45) [Found: *m/z* 416.1391. C₂₂H₂₆NO₃S (*M* + 1) requires *m/z*, 416.1428].

Method B. Ammonium molybdate(vi) tetrahydrate (2.16 g, 1.75 mmol) in H₂O₂ (30% in H₂O; 3.2 ml, 28 mmol) was added to a solution of the sulfide (2.68 g, 7.0 mmol) in EtOH (50 ml) at 0 °C. The cooling bath was removed and the reaction was followed by TLC (see method A). The mixture was diluted with Et₂O (200 ml) and water (200 ml). The organic phase was separated, washed with brine (200 ml), dried (MgSO₄), and filtered, and the filtrate was concentrated *in vacuo* to give a clear oil. Purification as described in method A gave the sulfone **7c** as a solid (2.18 g, 75%).

Dibenzyl *N*-(*tert*-butyldimethylsilyl)-D-aspartate **20**

NEt₃ (11.0 ml, 79.1 mmol) was added dropwise over a period of

1 h to a solution of TBDMSCl (5.7 g, 37.8 mmol), D-aspartic acid dibenzyl ester toluene-*p*-sulfonate¹⁷ **19** (16.2 g, 34.4 mmol) and DMAP (207 mg, 1.7 mmol) in CH₂Cl₂ (200 ml) under argon. The mixture was stirred for 16 h at rt and was then poured into saturated aq. NH₄Cl (200 ml), the organic layer was separated, washed (saturated aq. NaHCO₃), dried (Na₂SO₄), and evaporated *in vacuo* to give the *diester* **20** as an oil (14.4 g) which was used immediately in the next reaction. The ¹H NMR data agreed with those reported for the opposite enantiomer.^{4f}

(2*R*)-Benzyl *N*-(*tert*-butyldimethylsilyl)-4-oxoazetidine-2-carboxylate **22**

A solution of compound **20** (14.4 g) in dry Et₂O was cooled to 0 °C in an ice/salt-bath under argon. ⁴BuMgCl (20.0 ml, 40.0 mmol; 2 M in Et₂O) was added dropwise over a period of 40 min *via* a syringe pump. The mixture was kept at 0 °C for a further 1 h, the cooling bath was removed, and the solution was allowed to warm to rt and was stirred for 16 h. The mixture was recooled to 0 °C and saturated aq. NH₄Cl (20 ml) was added dropwise. The mixture was diluted with water (300 ml), the organic layer was separated, and the aqueous layer was extracted with Et₂O (2 × 200 ml). The combined organics were dried (MgSO₄) and the solvent was removed *in vacuo* to give a yellow oil (12.9 g) containing an equimolar amount of the *lactam* **22** and benzyl alcohol and which was used in the next reaction without further purification. The ¹H NMR spectrum for the *lactam* **22** agreed with that reported for the opposite enantiomer.^{4f}

(4*R*)-*N*-(*tert*-Butyldimethylsilyl)-4-(hydroxymethyl)azetidin-2-one **23**

A suspension of sodium borohydride (3.08 g, 81.5 mmol) in THF (150 ml) and a solution of lithium bromide (7.08 g, 81.5 mmol) in water (45 ml) were placed in the same dropping funnel and added dropwise over a period of 40 min to a solution of *lactam* **22** (12.9 crude, ~9.6 g *lactam*, ~30 mmol) in THF (150 ml) at such a rate that the internal temperature did not rise above 28 °C and the mixture was stirred for a further 40 min at rt. Saturated aq. NH₄Cl (75 ml) was added dropwise, the organic layer was separated, and the aqueous layer was extracted with ethyl acetate (2 × 150 ml). The combined organic extracts were dried (MgSO₄), and the solvent was removed *in vacuo* to give a yellow oil. Purification by column chromatography (silica gel; 30% ethyl acetate in petroleum spirit to 100% ethyl acetate) give the *alcohol* **23** (4.6 g, 62% from **19**) as an oil which solidified upon storage to give a low melting solid, [*a*] +30.2 (*c* 1, CHCl₃) {lit.,¹⁸ [(4*S*)-enantiomer] [*a*]_D -32.1 (*c* 1, CHCl₃) lit.,^{6b} [(4*S*)-enantiomer] [*a*]_D -31.5 (*c* 2.7, CH₂Cl₂)}. The spectral data for this compound agreed with those reported for the racemate⁴ⁿ and opposite enantiomer.¹⁸

(3*S*,4*R*)-*N*-(*tert*-Butyldimethylsilyl)-4-hydroxymethyl-3-methylazetidin-2-one **25**

Method A (LDA). A solution of compound **23** (650 mg, 3.0 mmol) in THF (20 ml) was added dropwise over a period of 20 min to a freshly prepared solution of LDA (6.7 mmol) in THF (10 ml) at -78 °C. The mixture was stirred at this temperature for 30 min, then methyl iodide (0.62 ml, 10.0 mmol) was added dropwise. The mixture was stirred for a further 2 h at -78 °C, MeOH (1.0 ml) was added dropwise followed by saturated aq. NH₄Cl (14 ml) and the mixture was allowed to warm to rt. The mixture was extracted with Et₂O (3 × 30 ml), the extract was dried (MgSO₄) and the solvent was removed *in vacuo* to give an oil. Column chromatography (silica gel; 50% ethyl acetate in petroleum spirit) gave the *alcohol* **25** (456 mg, 66%) as a solid, mp 63.5–65 °C (Found: C, 57.5; H, 10.1; N, 6.1. C₁₁H₂₃NO₂Si requires C, 57.6; H, 10.1; N, 6.1%); [*a*] +2.6 (*c* 1, CHCl₃); *v*_{max} (KBr) 3384s and 1702s cm⁻¹; ¹H NMR δ 0.25 (s, 3H), 0.26 (s,

3H), 0.96 (s, 9H), 1.31 (d, *J* 7.5, 3H), 1.70 [br s (exch), 1H], 3.05 (dq, *J* 2.4 and 7.5, 1H), 3.27 (ddd, *J* 2.4, 4.2 and 5.3, 1H), 3.68 (dd, *J* 5.3 and 11.5, 1H) and 3.76 (dd, *J* 4.2 and 11.5, 1H); ¹³C NMR δ -5.3, -5.1, 14.0, 18.8, 26.5, 49.1, 59.1, 64.5 and 177.2; *m/z* (CI) 230 (M⁺ + 1, 100%), 214 (15) and 174 (20).

Method B (*n*-BuLi). *n*-BuLi (1.2 M in hexane; 27.5 ml, 33.0 mmol) was added dropwise during 40 min to a solution of (4*R*)-*N*-(*tert*-butyldimethylsilyl)-4-(hydroxymethyl)azetidin-2-one **23** (3.50 g, 16.3 mmol) in THF (200 ml) at -78 °C. The mixture was stirred at this temperature for 45 min then methyl iodide (3.4 ml, 54.6 mmol) was added dropwise over a period of 15 min. The mixture was stirred for a further 2 h at -78 °C and then was worked up as described above. Column chromatography (silica gel; 45% ethyl acetate in petroleum spirit) gave the *alcohol* **25** (1.97 g, 53%) in addition to ~5% of (3*S*,4*R*)-4-(*tert*-butyldimethylsilyloxymethyl)-3-methylazetidin-2-one **26** (Found: C, 57.8; H, 10.2; N, 5.9%) [*a*] -44.8 (*c* 1, CHCl₃); *v*_{max}(KBr) 3199s and 1755s cm⁻¹; ¹H NMR δ -0.02 (s, 6H), 0.81 (s, 9H), 1.22 (d, *J* 7.5, 3H), 2.80 (ddq, *J* 1.0, 2.0 and 7.5, 1H), 3.27 (ddd, *J* 2.0, 4.9 and 5.8, 1H), 3.57 (dd, *J* 5.8 and 10.6, 1H), 3.67 (dd, *J* 4.9 and 10.6, 1H) and 6.50 (br s, 1H); ¹³C NMR δ -5.2, 13.1, 18.5, 26.0, 48.3, 57.5, 65.2 and 171.9; *m/z* (CI) 230 (M⁺ + 1, 100%), 185 (10), 158 (15) and 116 (15) [Found: M⁺ + 1, 230.1587. C₁₁H₂₄NO₂Si (M + 1) requires *m/z* 230.1598].

(2*R*,3*S*)-*N*-(*tert*-Butyldimethylsilyl)-3-methyl-4-oxoazetidine-2-carbaldehyde **8a**

Oxalyl dichloride (0.98 ml, 11.2 mmol) as a solution in CH₂Cl₂ (46.0 ml) was cooled to -78 °C, DMSO (1.49 ml, 21.05 mmol) as a solution in CH₂Cl₂ (6.0 ml) was added dropwise over a period of 10 min and the mixture was stirred for a further 15 min. A solution of compound **25** (1.90 g, 8.3 mmol) in CH₂Cl₂ (23.0 ml) was then added dropwise during 10 min and the mixture was stirred for 30 min before NEt₃ (5.35 ml, 38.4 mmol) was added. The mixture was stirred at -78 °C for 15 min then was allowed to warm to rt over a period of 1 h. Saturated aq. NH₄Cl (30 ml) was added and the mixture was poured into Et₂O (250 ml) and water (100 ml) and the organic layer was separated. The aqueous layer was extracted with Et₂O (3 × 80 ml) and the combined organics were washed with saturated aq. sodium chloride (2 × 150 ml), dried (Na₂SO₄), and evaporated *in vacuo* to give the *aldehyde* **8a** as a viscous oil (1.70 g, 90%), [*a*] +15.2 (*c* 1, CHCl₃); *v*_{max}(film) 1741 (br s cm⁻¹); ¹H NMR δ 0.13 (s, 3H), 0.29 (s, 3H), 0.96 (s, 9H), 1.42 (d, *J* 7.5, 3H), 3.24 (dq, *J* 2.7 and 7.5, 1H), 3.60 (dd, *J* 2.7 and 4.4, 1H) and 9.61 (d, *J* 4.4, 1H); ¹³C NMR δ -6.1, -6.0, 13.2, 18.1, 25.8, 49.4, 62.6, 188.4 and 199.0; *m/z* (CI) 228 (M⁺ + 1, 100%), 209 (35), 172 (30), 158 (15), 133 (20), 114 (40) and 71 (22) [Found: M⁺ + 1, 228.1433. C₁₁H₂₂NO₂Si (M + 1) requires *m/z* 228.1446].

General procedure for coupling compounds **7a–c** with *aldehyde* **8a**

Base (1 equiv.) was added dropwise to a solution of a compound **7a–c** in THF (~0.1 mM) at -78 °C and the mixture was stirred for 30 min. A solution of the *aldehyde* **8a** (1.1 equiv.) in THF (~0.2 mM) was added dropwise at -78 °C and the reaction mixture was stirred at this temperature before being allowed to warm to rt during 2 h. The reaction was quenched with saturated aq. NH₄Cl (20 ml) and the mixture was diluted with Et₂O (40 ml). The organic phase was separated and the aqueous phase was extracted with Et₂O (20 ml). The combined organic extracts were dried (MgSO₄), filtered, and concentrated *in vacuo* to give a yellow oil. Purification by column chromatography (silica gel; 50% Et₂O in petroleum spirit) gave the *lactam* **6a** as a clear oil. For reaction involving substrates **7a** and **7b**, material obtained in this way was of sufficient purity for characterisation. For reaction products from sulfone **7c**, final purification was effected after treatment with KF (see below).

(3*S*,4*S*)-*N*-[*tert*-Butyldimethylsilyl]-4-[(1*E*,3*E*,5*S*,6*S*)-6-methoxy-3,5-dimethyl-7-phenylhepta-1,3-dienyl]-3-methylazetididin-2-one 6a. ν_{\max} (film) 1747s cm^{-1} ; $^1\text{H NMR}$ δ 1.05 (d, *J* 6.8, 3H), 1.30 (d, *J* 7.5, 3H), 1.63 (d, *J* 1.1, 3H), 2.56–2.84 (m, 3H), 2.92 (dd, *J* 2.6 and 7.5, 1H), 3.15–3.20 (m, 1H), 3.23 (s, 3H), 3.62 (dd, *J* 2.6 and 9.1, 1H), 5.40 (d, *J* 9.9, 1H), 5.51 (dd, *J* 9.1 and 15.5, 1H), 6.20 (d, *J* 15.5, 1H) and 7.16–7.26 (m, 5H). $^{13}\text{C NMR}$ δ –5.92, –5.70, 12.4, 12.9, 15.8, 18.0, 25.9, 36.2, 37.8, 53.4, 58.3, 60.2, 86.6, 125.6, 127.3, 127.8, 129.0, 132.0, 136.0, 137.0, 139.0 and 176.0; *m/z* (CI) 428 ($M^+ + 1$, 100%), 412 (25), 294 (20) and 135 (40) [Found: $M^+ + 1$, 428.2989. $\text{C}_{28}\text{H}_{42}\text{NO}_2\text{Si}$ ($M + 1$) requires *m/z* 428.2993].

Diagnostic peaks for *E,Z*-diene: $^1\text{H NMR}$ δ 4.16 (dd, *J* 2.6 and 9.9, 1H) and 5.97 (d, *J* 11.3, 1H).

(3*S*,4*S*)-4-[(1*E*,3*E*,5*S*,6*S*)-6-Methoxy-3,5-dimethyl-7-phenylhepta-1,3-dienyl]-3-methylazetididin-2-one 31

KF (46.5 mg, 0.8 mmol) as a mixture in MeOH (6 ml) was added rapidly to a solution of the *N*-silyl lactam **6a** (280 mg, 0.7 mmol) in MeOH (15 ml) at 0 °C. The mixture was stirred at this temperature and monitored by TLC. After 4.5 h no starting material was present. Glacial acetic acid (40 μl) was added and the mixture was stirred for a further 10 min before being concentrated and the residue was purified by column chromatography (silica gel; 30–70% Et₂O in petroleum spirit) to give the lactam **31** as an oil (45% yield from sulfone **7c**, see text), ν_{\max} (film) 3202m and 1755s cm^{-1} ; $^1\text{H NMR}$ δ 1.04 (d, *J* 6.8, 3H), 1.34 (d, *J* 7.5, 3H), 1.65 (d, *J* 1.3, 1H), 2.56–2.96 (m, 4H), 3.21–3.24 (m, 1H), 3.23 (s, 3H), 3.78 [(after D₂O exch.) dd, *J* 2.6 and 8.0, 1H], 5.43 (d, *J* 10.4, 1H), 5.58 (dd, *J* 8.0 and 15.5, 1H), 5.90 [br s (D₂O exch.), 1H], 6.27 (d, *J* 15.5, 1H) and 7.11–7.31 (m, 5H); $^{13}\text{C NMR}$ δ 12.3, 15.8, 29.9, 36.2, 37.8, 53.5, 57.9, 58.2, 86.5, 125.5, 125.7, 127.8, 129.0, 131.9, 136.5, 137.0, 138.9 and 171.0; *m/z* (APCI) 314 ($M^+ + 1$, 100%), 180 (35) and 117 (60) [Found: $M^+ + 1$, 314.2103. $\text{C}_{22}\text{H}_{28}\text{NO}_2$ ($M + 1$) requires *m/z* 314.2086].

Diagnostic peaks for *E,Z*-diene: $^1\text{H NMR}$ δ 4.22 [ddd, *J* 0.9, 2.2 and 9.3; (after D₂O exch.) dd, *J* 2.2 and 9.3, 1H] and [6.01 (d, *J* 11.1, 1H)].

(3*S*,4*S*)-*N*-[*tert*-Butoxycarbonyl]-4-[(1*E*,3*E*,5*S*,6*S*)-6-methoxy-3,5-dimethyl-7-phenylhepta-1,3-dienyl]-3-methylazetididin-2-one 4

NEt₃ (0.19 ml, 1.4 mmol), a solution of (Boc)₂O (598 mg, 2.7 mmol) in CH₂Cl₂ (10 ml), and DMAP (167 mg, 1.4 mmol) were added to a solution of lactam **31** (430 mg, 1.4 mmol) in CH₂Cl₂ (10 ml). The mixture was stirred overnight at rt and concentrated. The $^1\text{H NMR}$ spectrum of the crude product (770 mg) showed only DMAP in addition to the lactam **4** (90–95% yield based on added DMAP). This material was either treated with lithium hydroxide (see below) to form *N*-Boc-ADDA or purified by column chromatography (silica gel; 30% Et₂O in petroleum spirit) to give the lactam **4** as an oil (340 mg, 60%), [α] –11.6 (*c* 1, CDCl₃); ν_{\max} (film) 1811s and 1718s cm^{-1} ; $^1\text{H NMR}$ δ 1.02 (d, *J* 6.8, 3H), 1.34 (d, *J* 7.5, 3H), 1.47 (s, 9H), 1.64 (d, *J* 1.3, 1H), 2.56–2.89 (m, 3H), 2.95 (dq, *J* 2.9 and 7.5, 1H), 3.15–3.22 (m, 1H), 3.22 (s, 3H), 4.02 (dd, *J* 2.6 and 8.3, 1H), 5.44 (d, *J* 12.0, 1H), 5.65 (dd, *J* 8.3 and 15.5, 1H), 6.33 (d, *J* 15.5, 1H) and 7.16–7.29 (m, 5H); $^{13}\text{C NMR}$ δ 11.3, 12.3, 16.1, 28.0, 36.6, 38.1, 51.6, 58.6, 61.4, 82.9, 86.8, 123.0, 126.0, 128.2, 129.4, 132.2, 137.2, 139.2, 147.9 and 168.4 (quat. aromatic not observed); *m/z* (APCI) 413 ($M^+ + 1$, 100%), 313 (50), 219 (60), 166 (60) and 150 (80).

Diagnostic peaks for *E,Z*-diene: $^1\text{H NMR}$ δ 4.49 (dd, *J* 2.9 and 9.7, 1H) and 6.09 (d, *J* 11.3, 1H).

{(2*S*,3*S*,4*E*,6*E*,8*S*,9*S*)-3-(*tert*-Butoxycarbonylamino)-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoyl}glycine methyl ester 5 (X = H, *n* = 0, R = Me)

NaN₃ (26 mg, 0.4 mmol) was added to a mixture of *N*-Boc

lactam **4** (70 mg, 0.17 mmol), glycine methyl ester hydrochloride (50 mg, 0.4 mmol) and NEt₃ (53 μl , 0.38 mmol) in DMF (3 ml). The mixture was stirred at rt for 5½ days, diluted with Et₂O, washed successively with water (3 × 25 ml) and aq. citric acid (10%; 2 × 25 ml), dried (MgSO₄), and filtered, and the filtrate was concentrated *in vacuo* to give a slightly yellow oil. Column chromatography (silica gel; 80% Et₂O in petroleum spirit) gave the starting material (7 mg, 10% recovery) in addition to the dipeptide **5** as a waxy solid (65 mg, 76%) (Found: C, 66.7; H, 8.7; N, 5.3. $\text{C}_{28}\text{H}_{42}\text{N}_2\text{O}_6$ requires C, 66.9; H, 8.4; N, 5.6%); [α] –13.0 (*c* 1.0, CHCl₃); ν_{\max} (KBr) 3310, 1762, 1688 and 1665 cm^{-1} ; $^1\text{H NMR}$ δ 1.02 (d, *J* 6.8, 3H), 1.24 (d, *J* 6.9, 3H), 1.44 (s, 9H), 1.62 (d, *J* 1.1, 3H), 2.56–2.70 (m, 3H), 2.81 (dd, *J* 4.5 and 13.8, 1H), 3.15–3.24 (m, 1H), 3.22 (s, 3H), 3.74 (s, 3H), 3.95–3.99 (m, 2H), 4.22 (m, 1H), 5.38 (d, *J* 9.9, 1H), 5.50 (dd, *J* 6.9 and 15.6, 1H), 5.80 (m, 1H), 6.20 (d, *J* 15.6, 1H), 6.34 (m, 1H) and 7.13–7.31 (m, 5H); $^{13}\text{C NMR}$ δ 12.7, 15.3, 16.3, 28.4, 36.5, 38.1, 41.0, 44.7, 52.3, 55.6, 58.5, 79.1, 86.9, 125.8, 125.9, 128.1, 129.3, 132.6, 135.6, 136.2, 139.4, 155.9, 170.1 and 175.1; *m/z* (CI) 503 (M^+ , 100%), 447 (20), 403 (45), 386 (25), 252 (100) and 135 (15) [Found $M^+ + 1$, 504.3191. $\text{C}_{28}\text{H}_{43}\text{N}_2\text{O}_6$ ($M + 1$) requires *m/z* 504.3199].

Reaction of samples of the *N*-Boc lactam **4** containing the 4*Z* isomer gave the corresponding dipeptide {(2*S*,3*S*,4*Z*,6*E*,8*S*,9*S*)-3-(*tert*-butoxycarbonylamino)-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoyl}glycine methyl ester **5** [α] –31.5 (*c* 0.66, CDCl₃); $^1\text{H NMR}$ δ 1.05 (d, *J* 6.8, 3H), 1.20 (d, *J* 7.1, 3H), 1.42 (s, 9H), 1.72 (d, *J* 1.1, 3H), 2.44–2.65 (m, 2H), 2.72 (dd, *J* 7.7 and 13.9, 1H), 2.86 (dd, *J* 4.6 and 13.9, 1H), 3.21–3.24 (m, 1H), 3.23 (s, 3H), 3.75 (s, 3H), 3.98 [d, *J* 5.3, 3H (D₂O exch.)], 3.98 (s, 3H), 4.73 (m, 1H), 5.26 (dd, *J* 9.7 and 11.7, 1H), 5.36 (d, *J* 9.7, 1H), 5.56 [m, 1H (D₂O exch.)], 5.92 (d, *J* 11.7, 1H), 6.15 [m, 1H (D₂O exch.)] and 7.18–7.27 (m, 5H); $^{13}\text{C NMR}$ δ 15.2, 16.3, 16.6, 28.4, 36.4, 38.1, 41.0, 45.5, 51.2, 52.4, 58.6, 79.0, 86.9, 125.9, 128.2, 129.4, 132.0, 134.6, 134.7, 139.5, 155.5, 170.2 and 174.9; *m/z* (CI) 503 (M^+ , 15%), 447 (15), 403 (100) and 252 (75) [Found: $M^+ + 1$, 504.3195. $\text{C}_{28}\text{H}_{43}\text{N}_2\text{O}_6$ ($M + 1$) requires *m/z* 504.3199].

(2*S*,3*S*,4*E*,6*E*,8*S*,9*S*)-3-(*tert*-Butoxycarbonylamino)-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoic acid *N*-Boc-ADDA

The crude mixture of *N*-Boc lactam **4** and DMAP prepared above was dissolved in THF (21 ml) and treated dropwise with aq. LiOH (4.1 ml; 1 M) at rt. The mixture was stirred at rt overnight. The THF was removed *in vacuo*, water (10 ml) was added and the solution acidified to pH 3.5–4.0 with 10% acetic acid, before being extracted with Et₂O (3 × 30 ml). The combined organics were dried (MgSO₄) and filtered, and the filtrate was concentrated *in vacuo* to give a yellow oil. Column chromatography (silica gel; 40% EtOAc in petroleum spirit) gave *N*-Boc-ADDA as an oil (510 mg, 86% yield for the two steps from lactam **31**). The spectral data for this compound agreed with those previously reported.^{8c,g,i,j}

(2*S*,3*S*,4*E*,6*E*,8*S*,9*S*)-3-Amino-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoic acid hydrochloride ADDA·HCl

A solution of *N*-Boc-ADDA (36 mg, 0.09 mmol) in EtOAc (1.5 ml) was treated with a 1.5 ml aliquot of a saturated solution of HCl in EtOAc. The mixture was stirred at rt for 4 h before being concentrated *in vacuo* to give ADDA·HCl as an oil (32 mg, 100%) whose spectra agreed with those reported previously^{8e} for the free amino acid: $^1\text{H NMR}$ (*d*₄-MeOH) δ 1.19 (d, *J* 6.8, 3H), 1.40 (d, *J* 7.1, 3H), 1.81 (s, 3H), 2.78–3.02 (m, 4H), 3.36–3.45 (m, 1H), 3.40 (s, 3H), 4.15 (m, 1H), 5.61–5.77 (m, 2H), 6.64 (d, *J* 15.3, 1H) and 7.28–7.41 (m, 5H); $^{13}\text{C NMR}$ (*d*₄-MeOH) δ 11.2, 13.0, 14.8, 35.7, 36.1, 37.3, 55.5, 57.2, 86.6, 119.0, 125.6, 127.7, 129.0, 131.8, 138.6, 138.9, 141.9 and 174.9.

(2S,3S,4E,6E,8S,9S)-3-Amino-9-methoxy-2,6,8-trimethyl-10-phenyldeca-4,6-dienoic acid ADDA

Method A. ADDA·HCl was dissolved in methanol and applied to a preparative RP-HPLC column (Econosil C18, I.D. 22 mm × 250 mm; flow rate 5.0 ml min⁻¹) and eluted with 6:4 methanol–water containing 5 mM ammonium formate. Collection of the eluent containing product and concentration gave free amino acid ADDA as a hygroscopic solid, $[\alpha]_{D} -38.9$ (c 0.375, EtOH); ¹H NMR (500 MHz; d₄-MeOH) δ 1.02 (d, J 6.8, 3H), 1.20 (d, J 7.2, 3H), 1.64 (s, 3H), 2.42 (ap. t, J 7.2, 1H), 2.62 (m, 1H), 2.68 (dd, J 7.3 and 14.0, 1H), 2.80 (dd, J 4.8 and 14.0, 1H), 3.24 (s, 3H), 3.21–3.27 (m, 1H), 3.68 (ap. t, J 8.4, 1H), 5.50 (dd, J 8.8 and 15.6, 1H), 5.54 (d, J 9.5, 1H), 6.41 (d, J 15.6, 1H) and 7.14–7.19 (m) and 7.22–7.26 (m, together 5H); ¹³C NMR (125 MHz; d₄-MeOH) δ 12.7, 16.1, 16.3, 37.7, 38.9, 44.8 br, 58.3, 58.7, 88.2, 122.7, 127.1, 129.2, 130.5, 133.5, 139.4, 140.5, 142.2 and 180.8; *m/z* (CI) 332 (M + H, 33%), 300 (15), 258 (10) and 135 (100).

Method B. ADDA·TFA (10 mg) was dissolved in aq. ammonia (5%; 1.0 ml) and freeze dried to give the free amino acid ADDA whose spectral details agreed with those reported above.

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